

Improving Seed Germination and Seedling Growth of Guava under Protected Conditions by Pre-Sowing Chemical and Hormonal Seed Treatments

Ajay Kumar Banyal^{1*}, Tarun Guleria² and Sanjeev Kumar Banyal¹

¹Associate Professor, Department of Fruit Science, College of Horticulture and Forestry, YS Parmar University of Horticulture and Forestry, Neri Hamirpur, (Himachal Pradesh), India.

²Ph.D. Scholar, Department of Fruit Science,

YS Parmar University of Horticulture and Forestry, Nauni Solan, (Himachal Pradesh), India.

(Corresponding author: Ajay Kumar Banyal*)

(Received 11 October 2021, Accepted 08 December, 2021)

(Published by Research Trend, Website: www.researchtrend.net)

ABSTRACT: Germination of the guava seeds is very poor, uneven and takes a long time because of the hard seed coats. Therefore, the present studies were carried out with the objective to reduce the difficulties in seed germination. Different pre-sowing seed dip treatments viz., GA₃ @ 400 and 600 ppm for 24 hours, HCl @ 2.5 and 5.0 % for 2 minutes, H₂SO₄ @ 2.5 and 5.0 % for 2 minutes, KNO₃ @ 0.5% and 1.0% for 24 hours, Cow urine @ 50 and 100 % purity for 24 hours and Control (direct sowing) in Completely Randomized Design with three replications were tried. It has been observed that among different pre-sowing treatments, when guava seeds were dipped in 1.0 % solution of KNO₃ for 24 hours, resulted in maximum (83.00%) guava seed germination. However, minimum time for seed germination (18.67 days), maximum mean daily germination (3.09), survival percent (91.67 %) and growth characteristics viz., seedling height, seedling diameter and number of leaves at 30, 60, 90 and 120 days after transplanting were observed, when guava seeds were dipped in 600 ppm solution of GA₃ for 24 hours.

Keywords: Guava, seed, KNO₃, GA₃, germination.

INTRODUCTION

Guava (*Psidium guajava* L.) is one of the most delicious fruit grown both in tropical and sub-tropical regions of the world and native to tropical America. It is also recognized as “poor man’s apple” owing to its cheaper cost and nutritional value (Dhaliwal and Singla, 2002). Guava has gained popularity all over world due to its nutritional importance and health benefits. It is a rich source of vitamins C, A, B₂ (riboflavin) and minerals like calcium, potassium, phosphorous and iron. It can be cultivated efficiently up to an elevation of 1500 m above mean sea level. However, temperature ranges between 20-30°C, well distributed rainfall ranges from 100 to 200 cm throughout the year, soils with good drainage, high levels of organic matter and pH ranges between 5.0 to 7.0 are ideal for successful guava cultivation (Yadava, 1996). In recent times, guava has witnessed a commendable growth in area expansion and its cultivation across the country. Guava occupies an area of 2,65,000 hectare with a production of 40,54,000 metric tonne in India (Anonymous, 2018a). In Himachal Pradesh, guava occupies an area of 2,292 hectares with annual production of 2,607 metric tonnes (Anonymous, 2018b). As the area under guava is increasing rapidly, the demand for vegetatively propagated plants has increased manifold and nurserymen has to produce a greater number of graftable seedling rootstock in shorter period. However, due

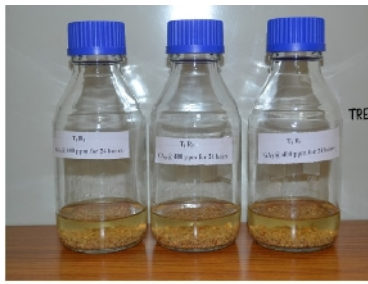
to the dormant cytological enzymes, presence of inhibitors and hard seed coat, there are more difficulties in the seed germination (Hejazi *et al.*, 2018). In recent years, attention has been shifted to the use of different chemicals and plant growth regulators to achieve higher, earliest seed germination and better seedling growth (Manthri and Bharad, 2017).

MATERIALS AND METHODS

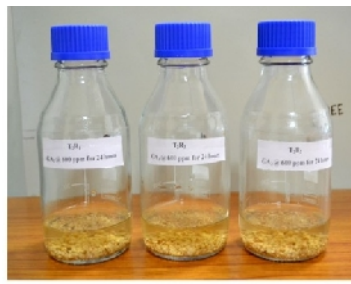
The present experiment was carried out in the nursery area of Department of Fruit Science, College of Horticulture and Forestry, Neri, Hamirpur, Himachal Pradesh during the academic year 2019-20. The experimental block of fruit nursery is located at an elevation of 618 m above mean sea level, between 31°41'49.98" North latitude and 72°28'02.21" East longitude. The experiment was laid out in Completely Randomized Design (CRD) with three replications using five grams (500 seeds approximately) of seeds. Guava seeds were treated with different pre-sowing treatments (Plate 1) viz., T₁: (GA₃ @ 400 ppm for 24 hours), T₂: (GA₃ @ 600 ppm for 24 hours), T₃: (HCl @ 2.5% for 2 minutes), T₄: (HCl @ 5.0% for 2 minutes), T₅: (H₂SO₄ @ 2.5% for 2 minutes), T₆: (H₂SO₄ @ 5.0% for 2 minutes), T₇: (KNO₃ @ 0.5% for 24 hours), T₈: (KNO₃ @ 1.0% for 24 hours), T₉: (Cow urine @ 50% for 24 hours), T₁₀: (Cow urine @ 100% for 24 hours) and T₁₁: (Control). The treated seeds were shade dried for 15 minutes and then sown in raised beds up to 0.5

cm depth under polyhouse conditions in first week of August. Guava seedlings were transplanted at 4-6 leaves stage after germination in polythene bags of 20 × 30 cm size and kept in polyhouse. Observations on seed germination (%), time taken for seed germination

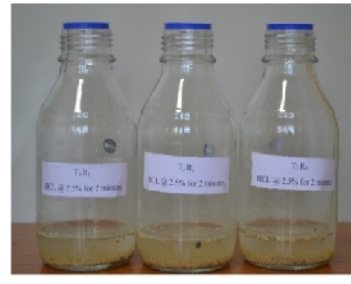
(days), mean daily germination (Samir *et al.*, 2015), survival and growth characteristics *viz.*, seedling height, seedling diameter and number of leaves at 30, 60, 90 and 120 days after transplanting were recorded.



T₁ (GA₃@ 400 ppm for 24 hours)



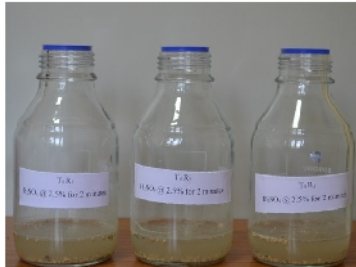
T₂ (GA₃@ 600 ppm for 24 hours)



T₃ (HCL@ 2.5% for 2 minutes)



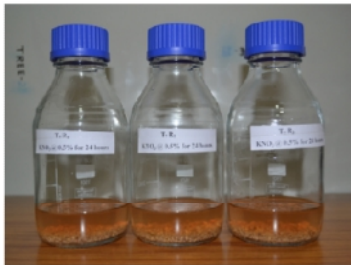
T₄ (HCL@ 5.0% for 2 minutes)



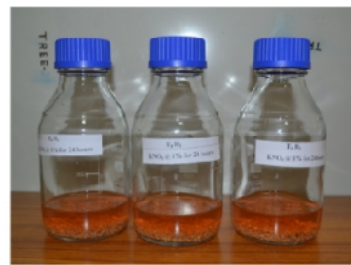
T₅ (H₂SO₄@ 2.5% for 2 minutes)



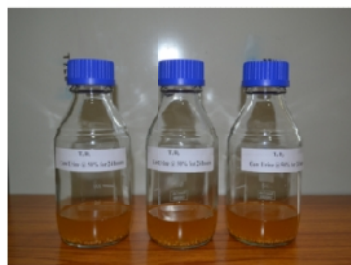
T₆ (H₂SO₄@ 5.0% for 2 minutes)



T₇ (KNO₃@ 0.5% for 24 hours)



T₈ (KNO₃@ 1.0% for 24 hours)



T₉ (Cow urine@ 50% for 24 hours)



T₁₀ (Cow urine@ 100% for 24 hours)



T₁₁ (Control)

Plate 1. Pre-sowing treatments in Guava seeds.

RESULT AND DISCUSSION

A. Seed germination (%)

The data pertaining to the effect of pre-sowing treatments are depicted in Table 1 showed a significant influence on percent seed germination. The maximum seed germination (83.00%) was observed in seeds which were dipped in 1.0 % solution of KNO₃ for 24 hours (T₈) and minimum seed germination (17.33%) was recorded with untreated control (T₁₁). The increase in germination might be due to the presence of nitrogenous compounds in KNO₃ which triggers the germination in soil by stimulating the pentose

phosphate pathway in seeds as it triggers the oxidation of NADPH to NADH. Nitrate present in KNO₃ can alter hormonal levels by inducing the expression of enzymes that catalyze the inactivation of abscisic acid and biosynthesis of gibberellins (Finch *et al.*, 2007). KNO₃ also removes the outer thin leathery seed coat, increase water absorption and led to enzyme activation and hydrolysis of food material (Aatla and Srihari 2013). These results are in conformity with those of Owino and Ouma (2011); Parab *et al.* (2017); Dwivedi *et al.* (2015) in papaya.

B. Time taken for seed germination (days)

The different pre-sowing treatments exerted a significant effect on the time taken for seed germination (Table 1). The minimum time for seed germination (18.67 days) was recorded in the seeds, which were treated with 600 ppm of GA₃ for 24 hours (T₂), while, maximum time taken for seed germination (29.67 days) was observed with control (T₁₁). The earliest seed germinations were attained due to gibberellins, which have a stimulatory effect in the formation of enzymes that induces faster radicle protrusion. Gibberellins acts on the embryo and causes denova synthesis of hydrolyzing enzymes particularly amylase and protease. The hydrolyzed food is utilized for growth of embryo and thereby enhanced the seed germination in guava (Kalyani *et al.*, 2014). These results are in harmony with those of Tandon *et al.* (2019) in Tamarind, Patel *et al.* (2016) in mango, Desai *et al.* (2017); Pratibha *et al.* (2015) in papaya.

C. Mean daily germination

The different pre-sowing treatments exhibited a marked effect on mean daily germination (Table 1). The highest mean daily germination (3.09) was recorded in the

seeds treated with 600 ppm of GA₃ for 24 hours (T₂). Whereas, lowest mean daily germination (0.39) was observed in the seeds of control (T₁₁). These findings are in conformity with those of Samir *et al.* (2015), where they found maximum mean daily germination when khirni seeds were presoaked in GA₃, because gibberellins resulted in higher germination percent and lower time for final germination.

D. Survival percent

The pre-sowing treatments showed a significant impact on the final survival of guava seedlings (Table 1). The maximum survival (91.67%) was obtained when guava seeds are treated with 600 ppm of GA₃ for 24 hours (T₂). However, minimum survival (25.67%) was recorded in control (T₁₁) treatment. The probable cause of higher survival percent of guava seedlings might be due to the easy root and shoot development and making the seedling stouter, withstanding the transplanting shock, resisting root diseases and leading to better growth (Palepad *et al.*, 2017). Our results are also in conformity with those of Barche *et al.* (2010) in papaya and Dinesh *et al.* (2019) in guava.

Table 1: Effect of pre-sowing treatments on percent seed germination, time taken for seed germination, mean daily germination and survival percent of guava seedlings.

Treatment	Percent seed germination (%)	Time taken for seed germination (days)	Mean daily germination	Survival percent (%)
T ₁ (GA ₃ @ 400 ppm for 24 hours)	50.67	25.00	1.52	75.00
T ₂ (GA ₃ @ 600 ppm for 24 hours)	75.33	18.67	3.09	91.67
T ₃ (HCL@ 2.5% for 2 minutes)	25.33	26.33	0.63	50.00
T ₄ (HCL@ 5.0% for 2 minutes)	30.33	25.67	0.77	70.00
T ₅ (H ₂ SO ₄ @ 2.5% for 2 minutes)	28.33	26.00	0.70	66.67
T ₆ (H ₂ SO ₄ @ 5.0% for 2 minutes)	35.66	25.33	0.94	72.67
T ₇ (KNO ₃ @ 0.5% for 24 hours)	68.33	23.00	2.38	83.33
T ₈ (KNO ₃ @ 1.0 % for 24 hours)	83.00	19.67	3.08	85.00
T ₉ (Cow urine@ 50% for 24 hours)	24.67	28.00	0.60	45.00
T ₁₀ (Cow urine@ 100% for 24 hours)	58.67	24.67	1.89	83.00
T ₁₁ (Control)	17.33	29.67	0.39	25.67
CD_{0.05}	3.22	2.54	0.14	10.90

E. Seedling height

The different pre-sowing treatments exerted a significant influence on seedlings height at 30, 60, 90 and 120 days after transplanting (Table 2 and Plate 2). The maximum seedling height (7.92 cm) was recorded with T₂ (GA₃@ 600 ppm for 24 hours) treatment at 30 days after transplanting. Whereas, minimum seedling height (1.94 cm) was registered in control (T₁₁).

At 60 days after transplanting, maximum seedling height (10.34 cm) was recorded with T₂ (GA₃@ 600 ppm for 24 hours) treatment and minimum seedling height (2.90 cm) was recorded with control (T₁₁).

At 90 days after transplanting, maximum seedling height (13.89 cm) was recorded when guava seeds were dipped in 600 ppm of GA₃ solution 24 hours (T₂), while, minimum seedling height (4.48 cm) was recorded with control (T₁₁), which was statistically at par with T₃ (5.56 cm) and T₅ (6.07 cm) treatment.

At 120 days after transplanting, maximum seedling height (18.89 cm) was recorded when guava seeds were dipped in 600 ppm of GA₃ solution 24 hours (T₂) and minimum seedling height (6.27 cm) was recorded in control (T₁₁).

The increase in seedling height might be due to the cell multiplication and elongation in the cambium tissue of the internodal region, as gibberellins apparently activates the metabolic processes or nullifies the effect of inhibitors on growth (Singh *et al.*, 1989). The gibberellin hormone increased the osmotic uptake of nutrients, causing cell multiplication and cell elongation in the cambium tissue of the internodal region; thus increased height of the plant (Patil *et al.*, 2018). These results are also in agreement with the findings of Biradar *et al.* (2005) in guava, Sheoran *et al.* (2018) in ber, Vasantha *et al.* (2014) in tamarind and Gurung *et al.* (2014) in passion fruit.

Table 2: Effect of pre-sowing treatments on seedling height at 30, 60, 90 and 120 days after transplanting of guava seedlings.

Treatment	Seedling height (cm)			
	30 days after transplanting	60 days after transplanting	90 days after transplanting	120 days after transplanting
T ₁ (GA ₃ @ 400 ppm for 24 hours)	6.55	8.71	10.24	13.90
T ₂ (GA ₃ @ 600 ppm for 24 hours)	7.92	10.34	13.89	18.89
T ₃ (HCL@ 2.5% for 2 minutes)	2.06	3.82	5.56	7.32
T ₄ (HCL@ 5.0% for 2 minutes)	4.03	5.61	9.45	12.52
T ₅ (H ₂ SO ₄ @ 2.5% for 2 minutes)	2.74	4.40	6.07	7.93
T ₆ (H ₂ SO ₄ @ 5.0% for 2 minutes)	2.98	5.63	9.82	12.69
T ₇ (KNO ₃ @ 0.5% for 24 hours)	4.90	7.30	10.82	14.73
T ₈ (KNO ₃ @ 1.0 % for 24 hours)	5.70	7.50	11.71	15.52
T ₉ (Cow urine@ 50% for 24 hours)	4.98	5.83	7.48	10.47
T ₁₀ (Cow urine@ 100% for 24 hours)	6.34	8.30	11.14	14.33
T ₁₁ (Control)	1.94	2.90	4.48	6.27
CD_{0.05}	2.00	2.36	2.64	2.94

F. Seedling diameter

The different pre-sowing treatments had a significant effect on seedling diameter at 30, 60, 90 and 120 days after transplanting (Table 3 and Plate 2). The maximum gain in seedling diameter (1.63 mm) was recorded in the plants raised from the seeds treated with 600 ppm of GA₃ for 24 hours (T₂), while, minimum growth in seedling diameter (0.92 mm) was recorded in control (T₁₁), at 30 days after transplanting.

At 60 days after transplanting, the maximum seedling diameter (2.11 mm) was obtained in the plants raised from the seeds treated with 600 ppm of GA₃ for 24 hours (T₂), however, minimum seedling diameter (1.23 mm) was observed in control (T₁₁).

At 90 days after transplanting, maximum growth in seedling diameter (2.49 mm) was obtained in the plants raised from the seeds treated with 600 ppm GA₃ for 24

hours (T₂), While, minimum seedling diameter (1.51 mm) was recorded in plants raised under control (T₁₁). Maximum growth in seedling diameter (3.17 mm) was attained when seeds were treated GA₃@ 600 ppm for 24 hours (T₂), however, minimum seedling diameter (1.74 mm) was recorded under control (T₁₁) at 120 days after transplanting.

The increase in seedling diameter might be due to fact that indigenous levels of gibberellins were increased with application of GA₃ that results in faster cell division, cell elongation and cell multiplication in the cambium tissue and reflects as increase in seedling diameter (Manthri and Bharad, 2017). Gibberellins might have increased the somatic uptake of nutrients, causing cell elongation and thus increasing height and diameter of the seedlings. These results are also in conformity with those of Rai *et al.* (2018) in khirni, Deb *et al.* (2010) in papaya and Al-Hawezy (2013) in loquat.

Table 3: Effect of pre-sowing treatments on seedling diameter at 30, 60, 90 and 120 days after transplanting of guava seedlings.

Treatment	Seedling diameter (mm)			
	30 days after transplanting	60 days after transplanting	90 days after transplanting	120 days after transplanting
T ₁ (GA ₃ @ 400 ppm for 24 hours)	1.49	1.97	2.27	2.53
T ₂ (GA ₃ @ 600 ppm for 24 hours)	1.63	2.11	2.49	3.17
T ₃ (HCL@ 2.5% for 2 minutes)	1.05	1.33	1.64	1.83
T ₄ (HCL@ 5.0% for 2 minutes)	1.20	1.38	1.66	2.14
T ₅ (H ₂ SO ₄ @ 2.5% for 2 minutes)	1.11	1.33	1.54	1.86
T ₆ (H ₂ SO ₄ @ 5.0% for 2 minutes)	1.46	1.86	2.22	2.52
T ₇ (KNO ₃ @ 0.5% for 24 hours)	1.52	1.89	2.30	2.86
T ₈ (KNO ₃ @ 1.0 % for 24 hours)	1.61	2.05	2.47	2.95
T ₉ (Cow urine@ 50% for 24 hours)	1.13	1.37	1.59	1.80
T ₁₀ (Cow urine@ 100% for 24 hours)	1.29	1.72	2.24	2.61
T ₁₁ (Control)	0.92	1.23	1.51	1.74
CD_{0.05}	0.30	0.35	0.38	0.33

G. Number of leaves

The pre-sowing treatments showed a significant effect on production of number of leaves at 30, 60, 90 and 120 days after transplanting (Table 4 and Plate 2). At 30 days after transplanting, maximum number of leaves (9.80) were formed in the plants raised from seed treated with GA₃@ 600 ppm for 24 hours (T₂), while,

minimum number of leaves (5.89) were formed in control (T₁₁).

At 60 days after transplanting, the maximum number of leaves (12.17) were obtained in the plants raised from seed treated with GA₃@ 600 ppm for 24 hours (T₂), which was statistically at similar with T₈ (11.70), T₇ (11.08) and T₁₀ (10.65) treatment. However, minimum number of leaves (6.73) were emerged in control (T₁₁)

and was statistically at par with T₃ (7.34), T₉ (7.58) and T₅ (8.13) treatment. While, maximum number of leaves (14.50) were obtained in the plants raised from seed treated with GA₃@ 600 ppm for 24 hours in (T₂) treatment and minimum number of leaves (8.10) were emerged in control (T₁₁), at 90 days after the transplanting.

Whereas, the maximum number of leaves (18.76) were formed the plants raised from seed treated with GA₃@ 600 ppm for 24 hours (T₂), while, minimum number of leaves (9.18) was obtained in control (T₁₁) at 120 days after transplanting.

Gibberellins moves into the shoot apex, increases cell division and cell growth apparently leading to increased development of young leaves (Salisbury and Ross, 1988). Therefore, the maximum number of leaves per seedlings in the present study with gibberellins may be due to acceleration of physiological processes and stimulatory action of gibberellins to form new leaves at a faster rate (Brijwal and Kumar 2013). These results are also in agreement with the findings of Jaiswal *et al.*, (2018) in kagzi lime and Singh and Maheswari (2017) in soursop.



T₁ (GA₃@ 400 ppm for 24 hours)



T₂ (GA₃@ 600 ppm for 24 hours)



T₃ (HCL@ 2.5% for 2 minutes)



T₄ (HCL@ 5.0% for 2 minutes)



T₅ (H₂SO₄@ 2.5% for 2 minutes)



T₆ (H₂SO₄@ 5.0% for 2 minutes)



T₇ (KNO₃@ 0.5% for 24 hours)



T₈ (KNO₃@ 1.0% for 24 hours)



T₉ (Cow urine@ 50% for 24 hours)



T₁₀ (Cow urine@ 100% for 24 hours)



T₁₁ (Control)

Plate 2. Vegetative growth of Guava seedlings after 120 days.

Table 4: Effect of pre-sowing treatments on number of leaves at 30, 60, 90 and 120 days after transplanting of guava seedlings.

Treatment	Number of leaves			
	30 days after transplanting	60 days after transplanting	90 days after transplanting	120 days after transplanting
T ₁ (GA ₃ @ 400 ppm for 24 hours)	7.78	9.57	11.77	14.76
T ₂ (GA ₃ @ 600 ppm for 24 hours)	9.80	12.17	14.50	18.76
T ₃ (HCL@ 2.5% for 2 minutes)	6.12	7.34	8.56	9.85
T ₄ (HCL@ 5.0% for 2 minutes)	6.43	8.95	10.86	12.70
T ₅ (H ₂ SO ₄ @ 2.5% for 2 minutes)	6.33	8.13	10.30	12.28
T ₆ (H ₂ SO ₄ @ 5.0% for 2 minutes)	6.45	9.60	11.37	14.58
T ₇ (KNO ₃ @ 0.5% for 24 hours)	8.83	11.08	13.17	16.42
T ₈ (KNO ₃ @ 1.0 % for 24 hours)	9.32	11.69	13.26	17.09
T ₉ (Cow urine@ 50% for 24 hours)	6.32	7.58	8.73	9.74
T ₁₀ (Cow urine@ 100% for 24 hours)	8.57	10.65	12.78	15.92
T ₁₁ (Control)	5.89	6.73	8.10	9.18
CD _{0.05}	1.70	1.87	2.46	2.76

CONCLUSION

The demand for the quality planting material of guava is increasing at faster pace across the whole country and it becomes hard to fulfill the requirement especially, seedling rootstock. Use of plant growth regulators/chemicals, that enhances the seed germination and their survival in fruit nurseries will boost the income of the growers. In the present study, it has been observed that guava seeds soaked in 600 ppm solution of GA₃ for 24 hours found most effective for earliest seed germination (18.67 days), improving mean daily germination (3.09), survival percentage (91.67%), seedling height, seedling diameter and number of leaves. However, guava seeds soaked in 1.0 percent solution of KNO₃ for 24 hours resulted in highest per cent seed germination (83.00%).

Acknowledgements. The author is thankful to the faculty members, Department of Fruit Science, College of Horticulture and Forestry Neri Hamirpur (H.P.) for providing all necessary facilities and support.

Conflict of Interest. None.

REFERENCES

Aatla, H. B. and Srihari, D. (2013). Influence of pre-sowing treatments on germination, growth and vigour of mango cv. Alphonso. *Asian Journal of Horticulture*, 8(1): 122-125.

Al-Hawezy, S. M. N. (2013). The role of the different concentrations of GA₃ on seed germination and seedling growth of loquat (*Eriobotrya japonica* L.). *Journal of Agriculture and Veterinary Science*, 4(5): 03-06.

Anonymous. (2018a). Area and production of horticulture crops: all India. *National Horticulture Board, Gurgaon*, (www.nhb.gov.in)

Anonymous. (2018b). Horticulture database, *Department of Horticulture, Government of Himachal Pradesh*, (www.hpagrinet.gov.in)

Barche, S., Kirad, K. and Singh, D. B. (2010). Response of seed treatment on germination, growth, survivability and economics of different cultivars of papaya (*Carica papaya* L.). *Acta Horticulturae*, 851: 279-283.

Biradar, S., Mukund, G. K. and Raghavendra, G. C. (2005). Studies on seed germination in guava cvs. Taiwan guava and Allahabad Safeda. *Karnataka Journal of Horticulture*, 1(3): 47-50.

Brijwal, M. and Kumar, R. (2013). Studies on the seed germination and subsequent seedling growth of guava (*Psidium guajava* L.). *Indian Journal of Agricultural Research*, 47(4): 347-352.

Deb, P., Das, A., Ghosh, S. K. and Suresh, C. P. (2010). Improvement of seed germination and seedling growth of papaya (*Carica papaya* L.) through different pre-sowing seed treatments. *Acta Horticulturae*, 851: 313-316.

Desai, A., Trivedi, A., Panchal, B. and Desai, V. (2017). Improvement of papaya seed germination by different growth regulator and growing media under net house condition. *International Journal of Current Microbiology and Applied Sciences*, 6(9): 828-834.

Dhaliwal, G. S. and Singla, R. (2002). Studies on the time of anthesis and dehiscence in different genotypes of guava in winter and rainy season crops. *Indian Journal of Horticulture*, 59(2): 157-161.

Dinesh, A., Padmapriya, S., Kavino, M., Raja, K. and Sujatha, K. B. (2019). Effect of different physical and chemical methods of seed treatment on germination and seedling growth attributes of guava (*Psidium guajava* L.). *Journal of Pharmacognosy and Phytochemistry*, 8(3): 4373-4377.

Dwivedi, D. H., Singh, S., Singh, N. and Kumar, P. (2015). Effect of plant bioregulator and chemical treatment on germination of papaya (*Carica papaya* L.) cv. Pusa Nanha. *Progressive Horticulture*, 47(2): 250-253.

Finch, W. E., Cadman, C. S., Toorop, P. E., Lynn, J. and Hilhorst, H. (2007). Seed dormancy release in *Arabidopsis* cvi by dry after ripening, low temperature, nitrate and light shows common quantitative patterns of gene expression directed by environmentally specific sensing. *Plant Journal*, 51(4): 60-78.

Gurung, N., Swamy, G. S. K., Sarkar, S. K. and Ubale, N. B. (2014). Effect of chemical and growth regulators on germination, vigour and growth of passion fruit (*Passiflora edulis* Sims.). *International Quarterly Journal of Life Sciences*, 9(1): 155-157.

Hejazi, Z., Sadat, M. I., Raghbi, M. G., Zerak, A. and Fitrat, K. (2018). Effect of different concentrations of GA₃, H₂O₂ and bleach solutions on seed germination of guava (*Psidium guajava* L.). *International Journal of Chemical Studies*, 6(5): 2679-2681.

Jaiswal, S. B., Nainwad, R.V., Supekar, S. J. and Mane, S. B. (2018). Effect of growth regulators and chemicals on growth of kagzi lime (*Citrus aurantifolia* Swingle.) seedlings. *International Journal of Current Microbiology and Applied Sciences*, 6: 940-944.

- Kalyani, M., Bharad, S. G. and Parameshwar, P. (2014). Effect of growth regulators on seed germination in guava. *International Journal on Biological Sciences*, 5(2): 81-91.
- Manthri, K. and Bharad, S. G. (2017). Effect of pre-sowing seed treatments on growth pattern of guava variety L-49. *International Journal of Chemical Studies*, 5(5): 1735-1740.
- Owino, D. O. and Ouma, G. (2011). Effect of potassium priming on papaya (*Carica papaya* var. Kamiya). *Journal of Animal and Plant Sciences*, 11(2): 1418-1423.
- Palepad, K. B., Bharad, S. G. and Bansode, G. S. (2017). Effect of seed treatments on germination, seedling vigour and growth rate of custard apple (*Annona squamosa*). *Journal of Pharmacognosy and Phytochemistry*, 6(5): 20-23.
- Parab, A. M., Mathad, J. C. and Malshe, K. V. (2017). Effect of pre-soaking chemicals on germination and subsequent seedling growth of papaya (*Carica papaya* L.) cv. Solo. *International Journal of Chemical Studies*, 5(4): 1812-1816.
- Patel, R. J., Ahlawat, T. R., Singh, A., Momin, S. K. and Chaudhri, G. (2016). Effect of pre-sowing treatments on stone germination and shoot growth of mango (*Mangifera indica* L.) seedlings. *International Journal of Agriculture Sciences*, 8(52): 2437-2440.
- Patil, H., Tank, R. V., Bennurmah, P. and Patel, M. (2018). Effect of seed treatment on germination and seedling growth of jamun (*Syzygium cuminii* L.). *International Journal of Current Microbiology and Applied Sciences*, 7(2): 2654-2659.
- Pratibha, C., Teja, T. and Krishna, P. M. (2015). Effect of chemical treatments on the germination and subsequent seedlings growth of papaya (*Carica papaya* L.) seeds cv. Pusa Nanha. *Journal of Agricultural Engineering and Food Technology*, 2(3): 189-191.
- Rai, R., Samir, M., Srivastava, R. and Uniyal, S. (2018). Improving seed germination and seedling traits by pre-sowing treatments in khirni (*Manilkara hexandra*). *Bulletin of Environment, Pharmacology and Life Sciences*, 7(4): 77-81.
- Samir, M., Ratna, R. R. and Prasad, B. (2015). Seed germination behavior as influenced by pre-sowing treatments in khirni. *Journal of Hill Agriculture*, 6(1): 132-135.
- Sheoran, V., Kumar, M., Yadav, S., Yadav, G. and Sharma, J. R. (2018). Effect of different seed scarification treatments on seed germination parameters of ber (*Ziziphus rotundifolia* Lamk.) under laboratory conditions. *International Journal of Current Microbiology and Applied Sciences*, 7(12): 1972-1980.
- Singh, M., Singh, G. H., Singh, L. N. and Singh, B. N. (1989). Effect of gibberellic acid on seed germination in mosambi (*Citrus sinensis* Obseek). *Haryana Journal of Horticultural Sciences*, 18(2): 29-33.
- Singh, S. J. and Maheswari, T. U. (2017). Influence of pre-sowing seed treatments on the performance of soursop (*Annona muricata* L.) seedlings. *Plant Archives* 17(2): 1215-1218.
- Tandon, K., Gurjar, P. K. S., Lekhi, R. and Soni, D. (2019). Effect of organic substances and plant growth regulators on seed germination and survival of tamarind (*Tamarindus indica* L.) seedlings. *International Journal of Current Microbiology and Applied Sciences*, 8(2): 2270-2274.
- Vasanth, P. T., Vijendrakumar, R.C., Guruprasad, T. R., Mahadevamma, M. and Santhosh, K. V. (2014). Studies on effect of growth regulators and biofertilizers on seed germination and seedling growth of tamarind (*Tamarindus indica* L.). *Plant Archives* 14(1): 155-160.
- Yadava, U. L. (1996). Guava (*Psidium guajava* L.): An exotic tree fruit with potential in the Southeastern United States. *Hort. Science*, 31(5): 789-794.

How to cite this article: Ajay Kumar Banyal, Tarun Guleria and Sanjeev Kumar Banyal (2022). Improving Seed Germination and Seedling Growth of Guava under Protected Conditions by Pre-Sowing Chemical and Hormonal Seed Treatments. *Biological Forum – An International Journal*, 14(1): 151-157.